

# **NWCA Field Demonstration Preparation**



Elizabeth Riley

# The Agenda Has Changed!



# Plan for the Field Day

- Rotating demonstration stations set up at 2 sites
  - SMALL groups (~10 people) rotate through each station within a site
  - Rotation proceeds 1-2-3-4-1
- Stations include:
  - VEG TEAM Stations
    - Vegetation Protocol
    - AA Station will go over assessment area establishment and USA-RAM AA metrics
  - AB TEAM Stations
    - Soil Protocol Station
    - Hydrology, Water Quality, Algae, Buffer Protocols and USA-RAM stressor metrics



# Plan for the Field Day

- Group A1 goes to Ginny Baker (NC DENR) and Elizabeth Riley (EPA) in Site A
- Group A2 goes to Mike Scozzafava (EPA) in Site A
- Group A3 goes to Cat McIntyre (MT NHP) in Site
- Group A4 goes to Gregg Serenbetz (EPA) and Regina Poeske (EPA) in Site C



# Plan for the Field Day

- Group B1 goes to Joanna Lemly (CO NHP) and Brian Gara (OH EPA) in Site B
- Group B2 goes to Chris Faulkner (EPA) in Site B
- Group B3 goes to Mary Anne Thiesing (EPA) in Site B
- Group B4 goes to Rick Savage (NC DENR) and Janet Nestlerode (ORD) in Site C



# LOGISTICS

Meet at 7:45 in hotel lobby to board bus

- Boxed lunches and water
  - Make sure you pay Tammy Taylor (\$10)

 Don't forget knee boots, rain gear, gloves and any other supplies

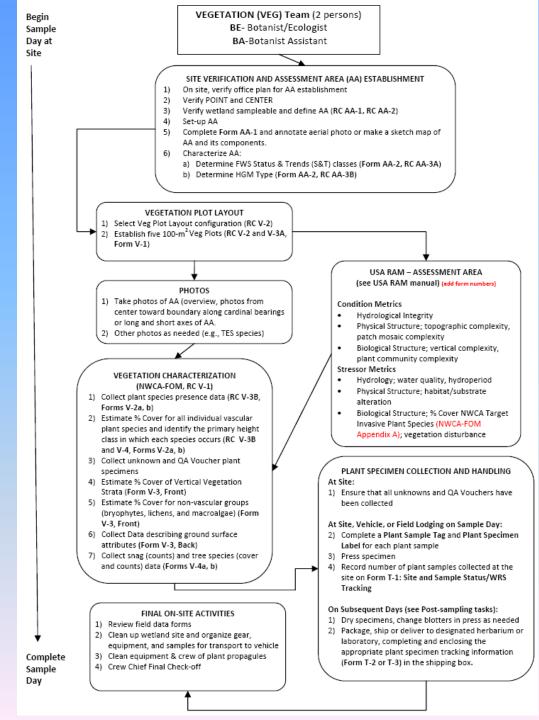
It is going to be COLD!



# PREPARATION FOR FIELD SAMPLING



# Vegetation Team Sampling Day





# Assessment Area/Buffer Team Sampling Day

ASSESSMENT AREA/BUFFER (AB) Team (2 persons)

AB1 – AB Member 1

AB2 – AB Member 2

### HYDROLOGY (Form H-1)

- 1) Identify Water Sources
- Determine and Rank Hydrology Stressors
- 3) Make Ditch Measurements
- Determine Presence of Hydrology Indicators (RC H-1)

WATER QUALITY (Form WQ-1)

- Complete Surface Water
   Characteristics Assessment
- Collect Water Sample and record associated data
- If desired, conduct Optional
   Water Quality Protocol with
   multi-probe meter
- Determine Maximum Depth and Surface Water Extent

### ALGAE (Form ALG-1, RC ALG-1)

- If any surface water is present, collect Chlorophyll-a sample. Collect Chl-a sample after collecting the Algal Toxin and Algae Taxonomic samples.
- 2) If both water and aquatic or emergent vegetation are present:
  - a) Collect epiphytic algae from vegetation
  - b) Collect phytoplankton from water
  - c) Composite a and b
  - d) Split this composite epiphyte-water subsample to form:
    - i) the Algal Toxin sample, and
    - ii) the Partial Algae Taxonomic sample
  - c) Collect substrate algae subsamples and combine with Partial Taxonomic sample to complete Algae Taxonomic sample
- 3) If water without vegetation is present, collect phytoplankton sample for:
  - a) the Algal Toxin Sample, and
  - b) the Partial Algae Taxonomic sample
  - Collect substrate algae subsamples and combine with Partial Taxonomic sample to complete Algae Taxonomic sample
- 4) If no water is present, collect Algae Taxonomic sample from substrate only
- Complete preparation of each sample by filtering or preserving

### SOILS - PROFILE DESCRIPTION (RC S-1A)

- Determine 4 Soil Pit locations (RC S-2A)
- Use a separate Form S-1 to record data for each pit
- At each Soil Pit, excavate to 60cm and record location, depth, and pit attribute information (Form S-1 (Front))
- Determine presence of Hydric Soil Indicators in the Soil
  Profile (RC S-5) and record on Form S-1 (Back)
- Describe soil profile at each Soil Pit by delineating horizons (RC S-2B and S-3) and by determining the following characteristics for each horizon. Record profile data on Form S-1 (Front):
  - a) H<sub>2</sub>S odor
- b) Horizon depth and horizon boundary abruptness
- c) Soil texture (RC S-4A)
- d) % Rock fragments and % Roots (RC S-3A)
- e) Soil matrix color (RC S-4B)
- Presence and type of redoximorphic or other features (RC S-3)
- g) % Surface area of distinct or prominent redox and other features (RC S-4B)
- h) Color of most evident redox or other feature (RC S-4B)
   Randomly select one Representative Soil Pit from the
- Excavate soil cores down to 125cm from the Representative Pit and describe the soil profile from 60cm down to 125 cm (Steps 3-6)

subset of pits with the most similar soils

### SOILS - SAMPLE COLLECTION IN REPRESENTATIVE PIT (RC S-1B)

- Once the Representative Pit has been identified, excavate as needed to collect four kinds of soil samples. Carefully bag and label each sample using prefilled labels and tags
- Collect the soil isotope (3 syringe cores) and the sediment enzyme (6 syringe cores) sample from surface soil layers at Representative Pit location
- Collect a Bulk Density (3 can cores) and a Chemistry/PSDA (1-2 liters) sample for each horizon > 8cm thick located in the top 60cm of the Representative Pit
  - Collect the Chemistry/PSDA sample for each horizon > 8cm thick located from 60 to 125cm deep in Representative Pit

### SOILS - DEPTH TO WATER IN SOIL PITS (Form S-1 (Back))

- After all data and all samples have been gathered at all four pits, collect water depth data at each pit.
- Backfill all 4 Soil Pits

### FINAL ON-SITE ACTIVITIES

- Complete preparation of unfinished water quality, algae, or soil samples
- 2) Record all sample types collected in the AA on Form T-1
- Review data form
- 4) Clean up site, gather equipment, and all samples for transport back
- 5) Clean equipment & crew of plant propagules
- 6) Crew Chief Final Check-off

NATURAL COVER AND STRESSORS

FOM - 12 Buffer Plots and AA CENTER Plot

A. In each of 3 Buffer Plots located along each

of 4 (N. W. E. S) 100m Buffer Transects, and in

the AA Center Plot, determine the presence

Natural Cover (Vegetation and Substrate)
 Residential and Urban Stressors

6) 22 Targeted Alien Species (presence only)

B. Record GPS coordinates of 3rd plot on each

USA RAM Metric 3: Stress to the Buffer Zone

1) While walking the 4 FOM Buffer Transects

(see above), record the presence and

2) Walk any other areas of the buffer zone

that aerial photos suggest might have

stressors to USA RAM Metric 3 Form.

anywhere along the transects.

severity of the buffer zone stressors (listed

on the USA RAM Metric 3 Form) that occur

potential stressors. If observed add these

3) Agriculture & Rural stressors

5) Hydrology Stressors

(USA RAM Manual)

4) Industrial Development Stressors

Plots are 100-m2 (10X10m) with visually

(Form B-1, RC B-1)

and abundance of:

estimated boundaries.

Begin

Sample Day at Site

e) Soil

Complete

Sample

Day



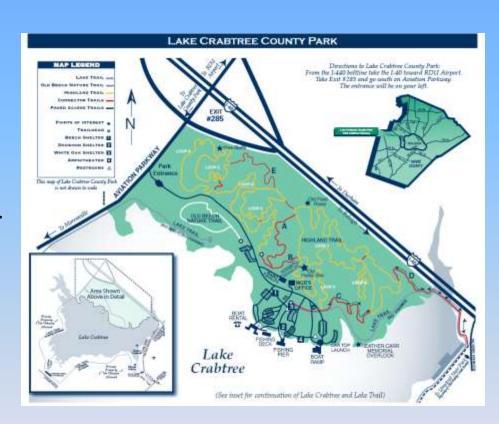
# Preparation for Field Sampling

- Before a field crew goes out to sample a site they must develop site packets containing:
  - appropriate maps,
  - aerial images,
  - contact information for the site (public/private),
  - photographs of the site,
  - copies of landowner permission forms,
  - all required permits to access and sample the site,
  - and any other specific requirements or instructions for accessing the site.



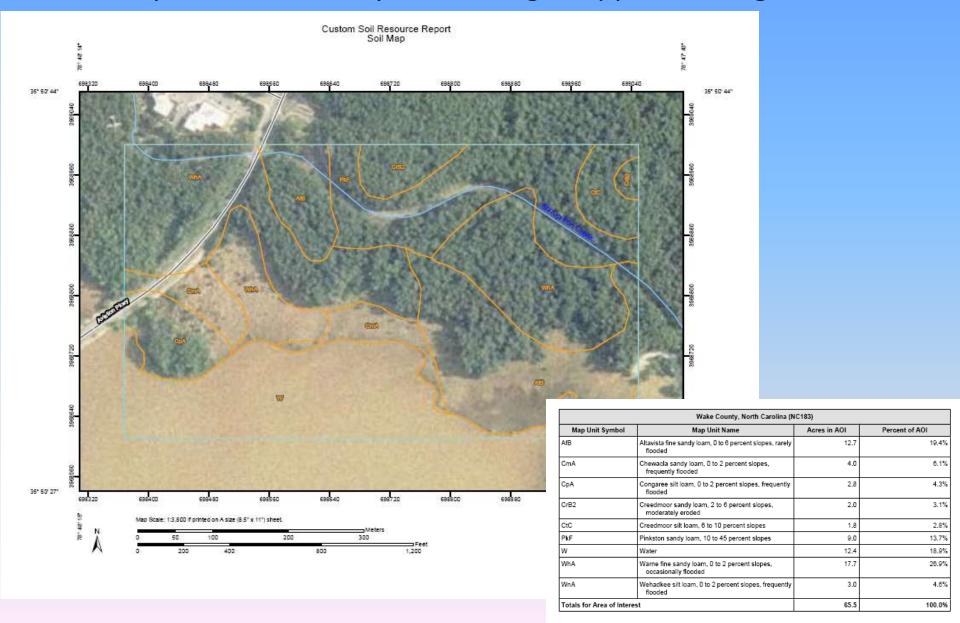
# Site Information

- Lake Crabtree County Park
  - 215-acre site adjacent to a 520-acre flood control lake
  - Currently used mainly for recreation purposes
  - Thank you to Rick Savage (NC DENR) for finding the site and obtaining permission to sample!



# Soils Map from Custom NRCS Soils Report

http://websoilsurvey.nrcs.usda.gov/app/HomePage.htm



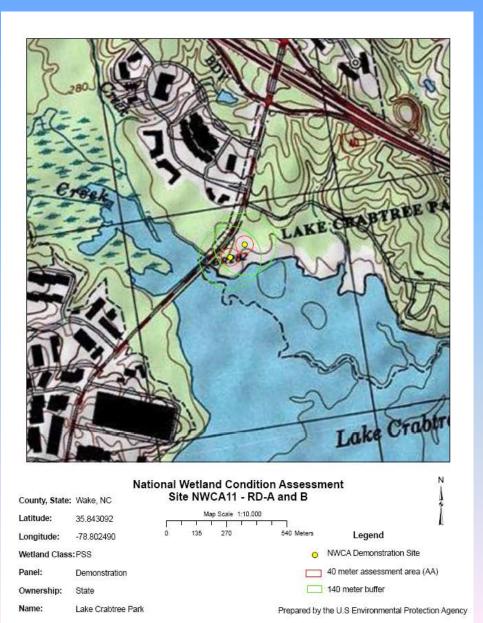


# Site Images



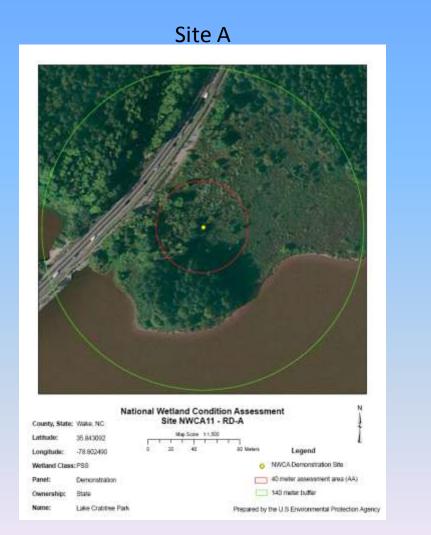


# **Topography Map**

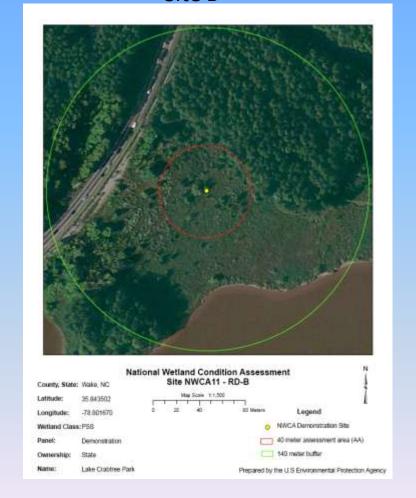




# Site Maps



Site B

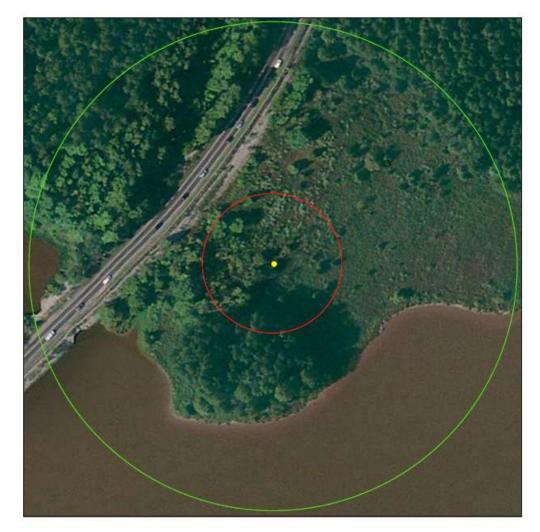


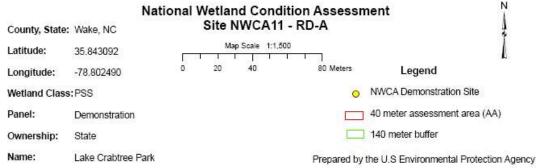


# ANNOTATING SITE MAPS AND USA-RAM BUFFER METRICS 1 & 2



# USA-RAM Metric 1: **51 – 75% Buffer**







## USA-RAM Metric 2

Line 1: 74 m

Line 2: 100m

Line 3: 100m

Line 4: 33m

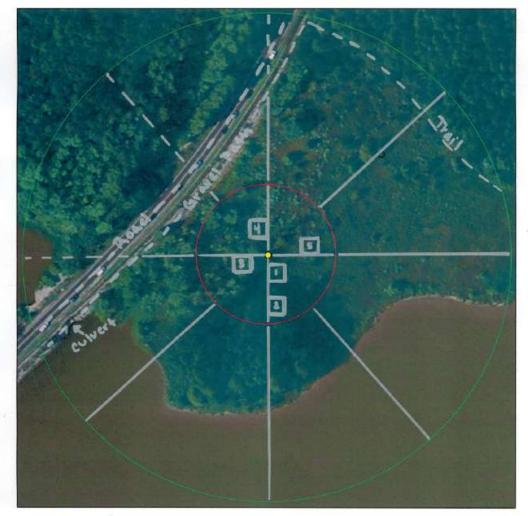
Line 5: 45m

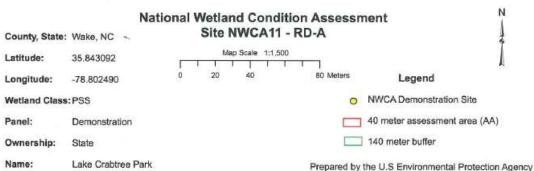
Line 6: 33m

Line 7: 45m

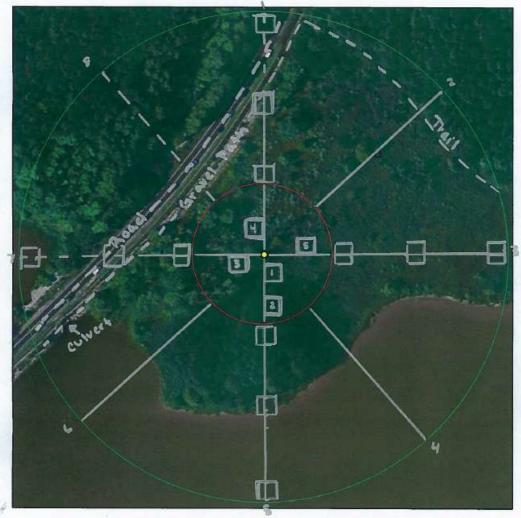
Line 8: 20m

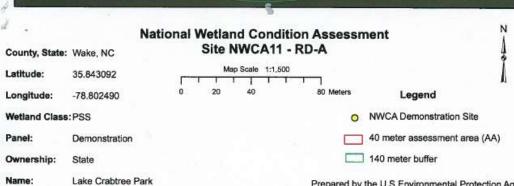
Mean Width = **56.25 meters** 











Prepared by the U.S Environmental Protection Agency

Name: